Date of Submission of the report: December 2001 **BARD Project Number:** IS 2686-96 Project Title: Modulation of the synthesis of the main preformed antifungal compound as a basis for the prevention of postharvest disease of C. gloeosporioides in avocado fruits **Investigators Institutions Principal Investigator (PI):** Volcani Center Dr. Dov Prusky **Co-Principal Investigator (Co-PI):** Dr. Noel T. Keen UC. Riverside Dr. John Browse Pullman University, Washington State Keywords. Fruit resistance, quiescent infections, Fruit diseases **Abbreviations commonly** used in the report, in alphabetical order: **Budget:** IS: \$ US: \$ Total: \$

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Modulation of the synthesis of the main preformed antifungal compound as a basis for the prevention of postharvest disease of *Colletotrichum gloeosporioides* in avocado fruits

The most important pathological factor limiting fruit life after harvest in subtropical fruits are quiescent infections of anthracnose caused by *Colletotrichum gloeosporioides*. Prusky and Keen elucidated the mechanism of resistance in avocado fruits to quiescent infections of *C. gloeosporioides* and determined that the major biocide involved is the preformed compound,1-acetoxy-2-hydroxy-4-oxo-heneicosa-13,15 diene.

Two possibilities exist for maintaining fungitoxic levels of antifungal compounds in the tissue of ripening fruits: (i). Prevention of catabolism (ii). Induction of synthesis. Previous work has demonstrated that increased fruit susceptibility after fruit harvest occurs through diene catabolism mediated by oxidation of the antifungal compound by the enzyme lipoxygenase. Levels of a non-specific inhibitor, epicatechin, in turn, regulate activity of lipoxygenase, present in the peel of unripe but not ripe fruit. In this proposal, we examined the possibility of exploiting induced synthesis of the antifungal compound for the study of the synthetic pathway.

The general objective of the present research was to study the mechanism of biosynthesis of natural antifungal compounds in order to regulate the process of resistance to postharvest diseases in ripening avocado fruits. The specific objectives of the research were:

- 1. To localize synthesis of the antifungal diene and modulate the process by biotic or a biotic elicitors.
- 2. To determine the relation between synthesis of the diene and accumulation in the peel and fruit resistance to decay
- 3. To characterize the biosynthetic pathway and the diene and the genes involved. The analysis of the antifungal compounds in avocado resulted in the detection of a new antifungal compound (E, Z, Z)-1-acetoxy-2-hydroxy-4-oxo-heneicosa-5, 12,15triene. This new compound was shown to inhibit spore germination of C. gloeosporioides similarly as the antifungal diene. We had localized one of the biosynthetic places of these antifungal compounds in specialized idioblast cells (oil cells) in the mesocarp that can be easily enhanced by elicitors as ethylene. Results have also suggested that the antifungal compounds can be "exported" from the mesocarp to the pericarp where its main activity takes place. The search for the biosynthesis of antifungal compounds and the genes involved took two directions i. direct search for specific genes involved in the synthesis of the diene and ii. Indirect selection of genes using the differential display library. We have cloned Δ^9 and Δ^{12} desaturase, a protein kinase and a elongase that their transcriptional activation is significantly enhanced during the enhanced synthesis of the antifungal diene. Although we are far away from a complete elucidation of the synthesis of the antifungal compound we have stepped forward determining some of the key steps that might be involved in its synthesis.

Achievements:

The main significant achievements of the present project are:

- 1. Elucidation of a new preformed antifungal compound in avocado fruits
- 2. Localization of the antifungal compounds and characterization of the mechanism of elicitation of the compounds by abiotic elicitors.
- 3. Cloning of the Δ^9 -stearoyl ACP desaturase, Δ^{12} -Fatty Acid Desaturase, Serine/threonine Kinase, and a fatty acid elongase from avocado fruit.
- 4. Characterization of the expression of Δ^9 -stearoyl ACP desaturase during biotic and abiotic stresses affecting the antifungal diene.

1. <u>Elucidation of a new preformed antifungal compound in avocado fruits</u>

A new compound was isolated from avocado, *Persea americana* Mill., idioblast cell oil. Its structure was determined as (E, Z, Z)-1-acetoxy-2-hydroxy-4-oxo-heneicosa-5, 12,15-triene. This new compound was shown to inhibit spore germination of the fungal pathogen *C. gloeosporioides*. The full characterization of other compounds that have been previously partially described is also reported.

2. Localization of the antifungal compounds and characterization of the mechanism of elicitation of the compounds by abiotic elicitors.

It has previously been demonstrated that exposure of whole avocado fruits cv. Fuerte to 40 μl/l ethylene for 3 h enhances the level of antifungal 1-acetoxy-2-hydroxy-4-oxo-heneicosa-12,15-diene in the fruit pericarp. Exposure of 1-2 mm slices of fruit pericarp and mesocarp to ethylene enhanced the level of the antifungal diene in the mesocarp only. Since most of the antifungal diene in the mesocarp is compartmentalized in idioblast the effect of ethylene was tested on isolated idioblast. Exposure of idioblasts to ethylene increased the level of antifungal diene twofold within 3 hs. This effect was temperature dependent. Three hours exposure of idioblast to ethylene at 35°C doubled the diene content compared to less than 50% increase after three hours at 20°C. Incubation of idioblast with [2-14°C] malonyl- CoA or [1-14°C] acetate in the presence of ethylene, showed the incorporation of the label

into a compound that co-chromatographed with the antifungal diene. The compound induced by ethylene and released from the cells was identified by NMR as the antifungal diene. The present report suggests that ethylene can induce the synthesis of the antifungal diene in idioblast and the export of this compound to the pericarp of the fruits.

3. Characterization of the expression of Δ^9 -stearoyl ACP desaturase during biotic and abiotic stresses affecting the antifungal diene.

The expression of the avocado homologue PAD1 encoding \Box^9 -stearoyl ACP desaturase was enhanced by multiple stimuli - inoculation with *Colletotrichum gloeosporioides*, exposure to ethylene or CO_2 , or low temperature (4 °C) and fruit wounding. This enhanced expression was correlated with an increase in the preformed antifungal (Z, Z)-1-acetoxy-2- hydroxy- 4-oxo- heneicosa-12, 15-diene. Treatments affecting up regulation of PAD1 increased as well the concentration of 18:2 fatty acid and the incorporation of C^{14} - linoleate to the antifungal diene. PAD1 expression was however down- regulated by light. Fruits with enhanced \Box^9 -stearoyl ACP desaturase expression were more resistant to *C. gloeosporioides*. It is suggested that the enhanced \Box^9 -stearoyl ACP desaturase expression which increases the concentration of unsaturated fatty acids, affects the synthesis of the This would result in the increase synthesis of the antifungal diene compound and the consequent enhanced resistance to fungal attack.

4. Cloning of the Δ 9-stearoyl ACP desaturase, Δ 12-Fatty Acid Desaturase, Serine/threonine Kinase, and a fatty acid elongase from avocado fruit.

As part of the effort to understand the possible mechanism of biosynthesis of the antifungal diene genes encoding for proteins at different points at the biosynthetic pathway were cloned. The characterization of these genes and the dynamics of their expression will be studied during the PhD student thesis of Xuejun Wang.

Cooperation:

A significant cooperation was kept since the early stages of the project through e-mails between all the participants. Dr. Fred Domergue, Post Doct working in Dr. Browse laboratory was in constant contact with the Israeli PI. As part of this interaction Dr. Prusky spent more than 2 months at the Institute of Biological

Chemistry. Washington State University at Pullman. During that period he was able to develop different aspects of the project in joint collaboration with Dr. Browse. Short visits took place by the PI in both Riverside California and Pullman, Washington, as part of the consulting and collaborations to solve specific problems.

List of publications

Reviewed papers:

Leikin, A. and Prusky, D. 1998.

Ethylene enhances the antifungal lipid content in idioblast cells from avocado mesocarp.

Phytochemistry 49: 2291-2298.

Domergue, F, Helms, G.H., Prusky, D., and Browse, J. 2000.

Antifungal compounds from idioblast cells isolated from avocado fruits. Phytochemistry 54:183-189.

Madi, L. and Prusky, D. 1999. Sequence of a cDNA clone encoding an avocado (*Persea americana*) Δ^9 stearoyl-acyl carrier protein desaturase. Plant Physiol. 121:1054.

Book Chapters:

- Leikin-Frenkel, A.I. and Prusky, D. 1997. Idioblast as a model system for the study of biosynthesis of lipids with antifungal properties in avocado fruits. In: Physiology, Biochemistry and Molecular Biology of Plant Lipids. ed. John P. Williams, Mobashsher U. Khan and Nora W. Lem Kluwer. Academic Pulbishers, Dordrecht.
- Prusky, D. 1998.Containment of fungal infection: preformed barriers.

 Postharvest Handling of Tropical Fruits. Australian Center for
 International Agricultural Research. 80:19-33
- Prusky, D., Ardi, R., Kobiler, I., Beno-Moualem, D., and Leikin, A. 1998.

 Mechanism of resistance of avocado fruits to *Colletotrichum*gloeosporioides attack. Postharvest Handling of Tropical Fruits.

 Australian Center for International Agricultural Research. 80:63-71.
- Prusky, D., Leikin-Frenkel, A., and Madi, L. 1999. Ethylene enhances the antifungal lipids content in idioblasts from avocado mesocarp. In Biology & Biotechnology of the Plant Hormone Ethylene II. Eds. A. K. Kanellis, C. Chang, H. Klee, A.B. Bleecker, J.C. Pech and D. Grierson. Kluwer Academic Publishers, 277-284.

Abstracts:

Leikin-Frenkel, A., Ardi, R., Kobiler, I. and Prusky D. 1997. treatments on the levels of antifungal compounds in oil cells of avocado mesocarp. Phytoparasitica 25:262

Effect of ethylene

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1. PUBLISHED PAPERS

2. SUBMITTED PAPERS

Plant Molecular Biology

Light and stress regulate Δ^9 -stearoyl ACP desaturase expression, fatty acids composition, antifungal diene level and resistance to fungal attack

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Abstract

The expression of the avocado homologue PAD1 encoding Δ ⁹-stearoyl ACP desaturase was enhanced by multiple stimuli - inoculation with *Colletotrichum gloeosporioides*, exposure to ethylene or CO₂, or low temperature (4 °C) and fruit wounding. This enhanced expression was correlated with an increase in the preformed antifungal (Z, Z)-1-acetoxy-2- hydroxy- 4-oxo- heneicosa-12,15-diene. Treatments affecting up regulation of PAD1 increased as well the concentration of 18:2 fatty acid and the incorporation of C¹⁴- linoleate to the antifungal diene. PAD1 expression was however down- regulated by light. Fruits with enhanced Δ ⁹-stearoyl ACP desaturase expression were more resistant to *C. gloeosporioides*. It is suggested that the enhanced Δ ⁹-stearoyl ACP desaturase expression that increases the concentration of unsaturated fatty acids, affects the synthesis of the antifungal diene compound and the consequent enhanced resistance to fungal attack.

Introduction

One of the primary products of fatty acid synthesis are palmitoyl-acyl-carrier (16:0-ACP) and D⁹ stearoyl -ACP (18:0-ACP). The soluble plastid enzyme, stearoyl-ACP desaturase (EC1.14.99.6), introduces the first double bond into stearoyl-ACP, between carbons 9 and 10, to produce oleoyl-ACP (18:1 Δ 9-ACP). Additional double bonds are introduced into oleic acid or palmitic acid by Δ ¹² desaturases after incorporation of the fatty acids into lipids. Δ 9 stearoyl-ACP desaturase activity is a key determinant of the overall level of fatty acid desaturation (Thompson *et al.*, 1991; Shanklin and Somerville, 1992).

In *Arabidopsis thaliana*, seven loci have been identified as fatty acid desaturase-encoding (Yadav *et al.*, 1993; McConn *et al.*, 1994; Ardi *et al.*, 1998). Of these, three loci- *fad3*, *fad7* and *fad8* are responsible for the production of trienoic fatty acids by unsaturation at the ω -3 position. Introduction of the *A. thaliana fad7* gene into tobacco under the control of a constitutive promoter resulted in a significant elevation of trienoic levels (Kodama *et al.*, 1994).

Plant desaturases respond to different environmental and physical stresses. Low temperature increases transcription of desaturase in maize, tobacco, arabidopsis and soybean (Cheesbrough, 1990; Miquel *et al.*, 1993; Somerville, 1995; Yong Moon *et*

al., 1995; Ishizaki-Nishizawa et~al., 1996). Light has also been reported as a stimulus that inducing desaturases (fad7) in A.~thaliana. In wheat, plastidic ω-3 desaturase gene expression rapidly increases when dark-adapted plants are transferred to white light (Horiguchi et~al., 1996).

Desaturase are also induced upon pathogen viroid inoculation in tomato plants (Gadea *et al.*, 1996) or by pathogen elicitors added to cultured parsley cells (Kirsch *et al.*, 1997). The function of antifungal compounds in modulating the level of avocado fruits resistance to *Colletotrichum gloeosporioides* leads us to try to understand the regulation and the possible stages involved in its biosynthesis (Prusky *et al.*, 1991; Prusky and Keen, 1995; Ardi *et al.*, 1998). The presence of a multiple double bond in the antifungal compound molecules ((Z, Z)-1-acetoxy-2-hydroxy-4-oxo-heneicosa-12, 15-diene and

(Z, Z, E)-1-acetoxy-2-hydroxy-4-oxo-heneicosa-5, 12,15-triene)) (Prusky and Keen, 1995; Domergue *et al.*, 2000) suggests that desaturation is an essential step in the biosynthesis of diene.

To initiate the study of diene biosynthesis regulation, we cloned stearoyl-ACP desaturase and tested its expression under conditions that enhance the level of antifungal diene (Ardi *et al.*, 1998; Leikin-Frenkel and Prusky, 1998) and the resistance of the avocado fruit to *Colletotrichum gloeosporioides*.

Materials and methods

cDNA libraries

Total RNA of ethylene- exposed and non exposed 10-month- old avocado fruits (

Persea americana Miller var. drymifolia (Schidl. and Cham.) S.F. Blake cv. 'Fuerte')

was isolated from the mesocarp using TRI REAGENT (Sigma St Louis, Mo). The

polyA+ RNA was isolated by using PolyATtract mRNA isolation systems (Promega).

Both ethylene exposed and non exposed cDNAs were synthesized by using ZAP

express cDNA synthesis kit (Stratagene, USA) and the libraries were packaged in the

ZAP express vector (Stratagene, USA) according to the manufacturer's instructions.

Biotic and abiotic treatments of the avocado fruits

All the treatments were applied on freshly harvested unripe cv. 'Fuerte' fruits.

Fruits were exposed to ethylene (40 ml/l), 30% CO₂, challenge inoculation with *Colletotrichum gloeosporioides* isolate Cg-14 and wounding as described by Ardi *et al.*, (1998). Low temperature included exposure of fruits to 4 °C (22 °C as a control) either in the dark (fruits were completely covered), or in the light (exposed to cool white fluorescent light at 140 mmol /m ² sec for up to 24 h).

The effect of shifting from dark to light was examined in fruits exposed for 18 h to darkness at 4 °C or 22 °C and then shifted to the light or maintained in the dark for another 6 h. Fruits exposed to the different treatments were sampled to determine Δ^9 stearoyl-ACP desaturase transcript level, diene level and decay development. These samples were frozen with liquid nitrogen and stored at -80 °C until use.

Fruit resistance to fungal attack and statistical analysis

A single-spore isolate of C. gloeosporioides (telemorph: Glomerella cingulata), isolate Cg-14, obtained from decayed avocado fruits was used for inoculation experiments (Yakoby et al., 2000). The fungus was maintained on Matur's medium (M3S media) at 20 °C. Inoculation was carried out on fruits following the different treatments by applying 10 ml of conidial suspension (1.5 x 10⁶ conidia /ml) at six inoculation spots on the longitudinal axis of the fruit, three on each side. Following inoculation, fruits were incubated at 22 °C, 90% RH for 24 h and then transferred to normal humidity conditions (70-80% RH) at 22 °C until fruit ripening and symptom development. Symptoms of decay on pericarp at the artificial inoculation sites, was denoted as dark spots and the lesion diameter expressed in mm. Symptoms of decay in naturally infected fruits was expressed as percentage of the infected area out of whole fruits as described earlier (Prusky et al., 1991) out of whole fruits as described earlier (Prusky et al., 1991). Inoculation experiments were repeated three times. Standard deviation of the means was calculated, and the differences between means were analyzed by analysis of variance. Presented results are from one representative experiment.

Northern blot analysis

Total RNA was isolated from avocado mesocarp, using a modified phenol/SDS method for plant RNA preparation (Ausbel *et al.*, 1992). Samples (0.5 g) were ground with a mortar and pestle under liquid nitrogen and were transferred to 6 ml grinding buffer pH 8.2 (0.18 M Tris-HCl; 0.09 M LiCl; 4.5 mM EDTA pH 8; 1 % SDS) and

mixed. To each sample, 6 ml phenol:chloroform were added and the sample was incubated at 50 °C for 20 min. The samples were subjected to phenol:chloroform extraction twice more and washed with 6 ml chloroform. The pellets were resuspended in 1 ml of water and precipitated twice with 2 M LiCl (final concentration), resuspended again in water and precipitated with sodium acetate/ethanol following washing with ethanol. RNA from each sample was denatured and separated (fifteen mg of per lane) in 1.1% formaldehyde agarose gels and transferred to Hybond-N membrane as previously described (Madi et al., 1993). The 1.8 kb EcoRI-XhoI fragment from pPAD1 (avocado cDNA) was prepared by random-primed synthesis. Probes were used in RNA blot analysis at a hybridization temperature of 42 °C in the presence of 50% formamide. Hybridizing PAD1 RNA was quantified with MacBAS v2.3 software (Fujifilm). To measure loading variations ethidium bromide -stained total RNA was quantified by photographing the gels using the Bio Imaging System (Dinco & Rhenium) and creating an image file using the Liscap software (Dinco & Rhenium). RNA in the different lanes was quantified using MacBAS v2.3 (Fujifilm).

Fatty acid analysis

Total fatty acid analysis was carried out according to Leikin-Frenkel and Brener, (1987) from 1 g of avocado pericarp or mesocarp. Fatty acid methyl esters were analyzed by gas liquid chromatography in Hewlett Packard equipment, built with a capillary column, apiezon-silica (Suopelco), and helium as the carrier gas. The temperature program was isothermic at 190 °C for 10 min, increasing at 1 °C per min for another 10 min, under these conditions all the fatty acids from miristate to eicosahexenoic, were separated according to their chain length and number of double bonds. The retention times were compared with those of true standards (Sigma) and the areas of the peaks were integrated. Quantification was also possible due to the incorporation of heptosadecanoic acid as an internal standard in all samples before the methylation step.

Antifungal diene extraction

Avocado pericarp or mesocarp (1-2 g,1- 2-mm deep slices) were homogenized in 95% ethanol in an Omni-Mixer (Sorvall; Dupont Co., Newton, CT.) at full speed for 3 min. The ethanol extract was dried in a rotary evaporator at 40 °C, brought to 10 ml in

distilled water, and the organic phase was extracted by fractionation with dichloromethane. Following two extractions, the organic phases were pooled, dried with anhydrous MgSO₄ (Riedel-deHaen, Seelze, Germany), and evaporated to dryness. Samples were dissolved in 1 ml of ethanol (analytical grade; BioLab, Jerusalem, Israel) and analyzed by high-performance liquid chromatography (Prusky *et al.*, 1991). Each experiment was run in four replicates. The diene level was expressed as mg / g of fresh weight (FW). Standard deviation of the means was calculated, and the differences between means were analyzed by analysis of variance.

Incorporation of C^{14} *linoeate.*

Avocado pericarp (or mesocarp) 1mm thick, was sliced into small pieces (3x1x1mm). Three grams of tissue per sample was incubated with labeled (1-¹⁴C) linoleate (NEN, Boston Ma. USA) in 5 ml of 50 mM phosphate buffer pH 7.2 for different length of time (1-24 h) at room temperature. Addition of equal volume of ethanol ended the reaction. At zero time controls the ethanol were added with the labeled linoleate. The non-incorporated linoleate was washed twice with phosphate buffer. Diene extraction was done as described above, and separated by RP-HPLC. The antifungal diene was collected as described before and radioactivity was measured in the presence of Ultima Gold (Packard Inst. Co. Meriden, Ct. USA) in a liquid sentillation counter.

Results

Cloning of a Δ^9 stearoyl-ACP desaturase from avocado

cDNA libraries constructed from ethylene- exposed and nonexposed avocado fruits were screened, using the full - length cDNA clone pMRCD9 of *Ricinus communis* as a probe. As a result of screening $4x10^4$ clones in both libraries, 23 putative clones from the ethylene- exposed cDNA, vs. only five putative clones from the nonexposed cDNA were isolated. Four clones were sequenced and three shared homology to Δ^9 stearoyl-ACP desaturase. One clone, PAD1, a 1818-bp showed 77% homology to the DNA sequence of pRCD1 from *R. communis*. PAD1 contains a 1188-bp open reading frame and encodes a 396-amino- acid protein of 45.4 kDa with 89% amino acid homology and 84% identity to the castor clone pRCD1. PAD1 also shares considerable homology with known Δ^9 stearoyl-ACP desaturases (Thopson *et al.*, 1991; Arondel *et al.*, 1992; Sato *et al.*, 1992; Taylor *et al.*, 1992; Cahoon *et al.*, 1994). The deduced amino- acid sequence contains two regions of sequence identity to the

two peptides (DETGASP and DYADILE) identified by direct N-terminal sequencing of tryptic fragments derived from the purified avocado stearoyl ACP desaturase identified by Shanklin and Somerville (1992). The alignment of the deduced amino acid sequence of the avocado clone, PAD1 (AF116861), with the desaturases of castor (M59857), cucumber (M59858), *Helianthus* (U91339) and *Brassica* (X74782) (Figure 1) shows the high sequence similarity between desaturases of five distantly related families, suggesting that the sequences are highly conserved among higher plants.

Induction of PAD1 transcript and the antifungal diene by different stimuli.

Inoculation of fruits with C. gloeosporioides or exposure to ethylene (40 μ l/l), 30% CO_2 or wounding all showed increases of 25% to 50% in the mesocarp antifungal diene over control fruit (Table 1). Diene level in the mesocarp of ethylene-treated fruits was elevated after 3 h of exposure. Incorporation of C^{14} -linoleate into the antifungal diene following exposure to ethylene showed increased 3 h but the level of the C^{14} labeled diene declined 2 h later (Figure 2).

Analysis of the total RNA extracted from the mesocarp revealed that PAD1 transcript increased significantly in all treatments which induced diene level, PAD1 transcript was most abundant after 24 h of treatment, whereas some induction in the control fruits was also observed at 24 to 48 h (Figure 3).

Effect of temperature and light conditions on PAD1 transcription, fatty acids composition and the antifungal diene levels

Exposure of avocado fruit for 6 h to 4 °C enhanced PAD1 transcription more than at 22 °C, no matter if the fruits were exposed to light or darkness; however dark treatments induced higher expression than light treatments (Figure 4). If fruits were exposed to a longer darkness period (18 h) at 4 °C or at 22 °C PAD1 expression was higher in fruits kept in the dark, no matter the temperature, than in the tissue of fruits transferred to light for another 6 h. (Figure 5).

Fatty acid composition (Figure 6A) in mesocarp of fruits exposed to 4 °C at dark show that the content of 18:2 and the antifungal diene levels were higher by 33% and 45% respectively than in fruit exposed 22 °C (Figure 6B).

Effect of temperatures and light/dark conditions on decay development

To examine possible involvement of Δ^9 stearoyl-ACP desaturase in defense response of avocado to *C. gloeosporioides*, we followed the decay development in fruits with elevated or reduced D⁹ desaturase expression. Decay development in the artificially (Figure 7A)or naturally (Figure 7B) infected fruits was significantly higher when fruit exposed to light at 22 °C Whereas when fruits exposed to dark at 22 °C or to light at 4 °C the decay was significantly reduced. The lowest level of decay was always observed when fruits were exposed to 4 °C in dark (Figure 7A and B).

Discussion

The preformed antifungal (Z, Z)-1-acetoxy-2- hydroxy- 4-oxo- heneicosa-12, 15diene is involved in avocado resistance to C. gloeosporioides. Several elicitors have been reported to induce the increase of this compound (Prusky and Keen, 1995; Ardi et al., 1998; Leikin-Frenkel and Prusky 1998) however the biosynthetic pathway involved was never described. We have assumed that synthesis of the diene involves extension of a linoleic acid starting compound by conventional two-carbon elongation. As a first stage to determine the biosynthetic pathway involved in the synthesis of the antifungal diene we have found that Δ^9 stearoyl-ACP desaturase cloned from avocado fruits is significantly expressed by similar elicitors that induce the antifungal diene, ethylene (40 µg/l), CO2 (30%) wounding, C. gloeosporioides inoculation and cold treatment (4 °C). Elevated transcription of desaturases (ω -3, Δ ² and Δ 9) at low temperature had been reported in plants (Miguel et al., 1993; Somerville, 1995; Young Moon et al., 1995; Ishizaki-Nishizawa et al., 1996) and was implied to be required for resistance to chilling by altering the lipids composition in the membrane (Miquel et al., 1993). In avocado, the increase in Δ 9 stearoyl-ACP desaturase transcripts level in fruits exposed to

 4 0 C compared to fruits exposed to 2 0 C, suggest that avocado fruit responds to low temperature in the same manner like other plants by increasing the 9 stearoyl-ACP desaturase transcription level and the unsaturated fatty acids level (mainly linoleic acid). However, no report has been published on induction of 9 stearoyl-ACP desaturase in correlate with the increase of antifungal diene synthesis and its effect on fungal resistance.

Another elicitor that was found to activate the PAD1 is storage under darkness. In *A. thaliana FAD7* expression and the wheat plastidic ω -3 desaturase transcripts

rapidly increased when dark-adapted plants were transferred to white light (Horiguchi *et al.*, 1996). In avocado however, Δ 9 stearoyl-ACP desaturase expression level increased in fruits exposed to darkness compared to fruits exposed to light. The effect of light on the transcription level is more pronounced when fruits exposed to darkness were transferred to light. In this case the level of transcription was markedly reduced independently of the temperature that fruits were exposed.

The concurrent increase in Δ^9 stearoyl-ACP desaturase transcription, 18:2 content, and the incorporation to C^{14} into the antifungal diene suggest the Δ^9 - stearoyl-ACP desaturase is involved in the diene biosynthesis probably by increasing the level of the diene precursors, that can incorporate into the antifungal diene. This increase in diene level significantly affect fruit susceptibility to C. gloeosporioides attack, as observed in fruits exposed to 4 °C at darkness. Alternatively induction of desaturases (Gaeda et al., 1996; kirsch et al., 1997) and lipids desaturation was suggested to be an early component of the complex of defense response (Kirsch et al.,1997). Acquired resistance to fungal attack mediated by unsaturated lipids as a result of increased activity of desaturase had been reported so far only by Wang et al., (1999), where it was shown that expression of Λ^9 stearoyl-ACP desaturase of yeasts in tomato plants enhanced its resistance to powdery mildew by elevating desaturated fatty acids. Cohen et al., (1991) had reported that external treatment of plant leaves with unsaturated fatty acids induced systemic resistance in potato plant against Phytophthora infestans. Unsaturated fatty acids can transform to lipids peroxides that can acts as antimicrobial compounds Wang et al., (1999), or by stimulating the signal transduction pathway via octadecanoid pathway to signal molecules such as jasmonate (Vick and Zimmermann, 1984; Tzeng and DeVay, 1993). Modulation of resistance by ROS induced in unripe avocado fruits by fungal infection was suggested by Beno-Moualem and Prusky (2000).

Present results indicate that elicitors activate Δ^9 stearoyl-ACP desaturase transcript expression, linoleic acid and antifungal diene content (Prusky *et al.*, 1991; Prusky and Keen , 1995). Although our data does not directly support the involvement of desaturase in diene synthesis, the data presented suggest that desaturase is an important step in diene synthesis.

References

Ardi, R., Kobiler, I., Jacoby, B., Keen, N.T. and Prusky, D. 1998. Involvement of epicatechin biosynthesis in the activation of the mechanism of resistance of avocado fruits to *Colletotrichum gloeosporioides*. Physiol. Mol. Plant Pathol. 53: 269-28.

Arondel, V., Lemieux, B., Hwang, I., Gibson, S., Goodman, H.M., and Somerville, C.R. 1992. Maped based cloning of a gene controlling omega-3 fatty acid desaturation in arabidopsis. Science 258: 1353-1354.

Ausbel, M.F., Brent, R.R., Kingston, E.R., Moore, D.D., Seidman, G.J., Smith, A.J., Struhl, K., 1992. In Current Protocol in Molecular Biology. John Wiley and Sons.

Beno-Moualem, D. and Prusky, D. 2000. Signaling processes involved in the activation of defense mechanism of avocado fruits against *Colletotrichum gloeosporioides*. Phytopathology 90: 553-559.

Berberich, T., Harada, M., Sugawara, K., Kodama, H., Hiroaki, Iba, K. and Kusano, T. 1998. Two maize genes encoding ω-3 fatty acid desaturase and their differential expression to temperature. Plant Mol. Biol. 36:297-306.

Cahoon, E.B., Becker, C.K., Shanklin, J., Ohlrogge, J.B. 1994. cDNAs for isoform of the Δ ⁹– stearoyl–acyl carrier protein desaturase from *Thunbergia alata* endosperm. Plant Physiol. 106: 807-808.

Cheesbrough, T.M. 1990. Regulation of stearoyl-acyl carrier protein desaturase. Plant Physiol. 93: 555-559.

Cohen, Y., Gisi, U. and Mosinger, E. 1991. Systemic resistance of potato plants against phytophthora infestans induced by unsaturated fatty acids. Physiol. Mol. Plant Pathol. 38: 255-263.

Domergue, F., Gregory, L.H., Prusky, D. and Browse, J. 2000. Antifungal compounds from idioblast cells isolated from avocado fruits. Phytochemistry 54: 183-189.

Gadea, J., Mayda, M.E., Conejer, V., Vera, P. 1996. Characterization of defense-related genes ectopically expressed in viroid-infected tomato plants. Mol. Plant Microb. Interact. 9:409-415.

Horiguchi, G., Iwakawa, H., Kodama, H., Kawkami, N., Nishimura, M. and Iba, K. 1996. Expression of a gene for plastid ω -3 fatty acid desaturase and changes in lipid and fatty acid compositions in light – and dark grown wheat leaves. Physiol. plant 96:275-283.

Ishizaki-Nishizawa, O., Fujjii, T., Azuma, M., Sekiguchi, K., Murata, N., Ohtani, T., Toguri, T. 1996. Low-temperature resistance of higher plants is significantly enhanced by a non-specific cyanobacterial desaturase. Nature Biotechnol. 14: 1003-1006.

Kirsch, C., Hahlbrock, K., Somssich, I.E. 1997. Rapid and transient induction of parsley microsomal delta 12 fatty acid desaturase mRNA by fungal elicitor. Plant Physiol. 115:283-289.

Kodama, H., Hamada, T., Horiguchi, G., Nishimura, M., and Iba, K. 1994. Genetic enhancement of cold tolerance by expression of a gene for chloroplast ω -3 fatty acid desaturase in transgenic tobacco. Plant Physiol. 105: 601-605.

Leikin-Frenkel, A. and Prusky, D. 1998. Ethylene enhances the antifungal lipids contents idioblasts from avocado mesocarp. Phytochemistry 49: 2291-2298.

Leikin-Frenkel, A. and Brener, R.R. 1987. Cholesterol induced microsomal changes modulate desaturase activities. Biochem. Biophys. Acta 922: 294-303.

Madi, L., McBride, S., Baily, L. and Ebbole, D.J. 1993. Mutants of Neurospora *crassa* that alter gene expression and conidia development. Proc. Natl. Acad. Sci. USA 91:6226-6230.

Madi, L. and Prusky, D. 1999. Sequence of a cDNA clone encoding an avocado (Persea americana) Δ^9 stearoyl-acyl carrier protein desaturase. Plant Physiol. 121:1054.

McConn, M., Hugly, S., Browse, J. and Sommerville, C. 1994. A mutation at the *fad8* locus of Arabidopsis identifieds a second chloroplast ω -3 desaturase. Plant Physiol. 106: 1609-1614.

Miquel, M., James, D., J.R., Dooner, H. and Browse, J. 1993. Arabidopsis requires polyunsaturated lipids for low-temperature survival. Proc. Nat. Acad. USA 90: 6208-6212.

Nishida, I., Beppu, T., Matsuo, T., Murata, N. 1992. Nucleotide sequence of a cDNA encoding a precursor to stearoyl –(acyl-carrier-protein) desaturase from spinach, *Spinacia oleracea*. Plant Mol. Biol. 19: 711 -713.

Nishiuchi, T., Nishimuri, M., Arondei, V., Iba, K. 1994. Genomic nucleotide sequence of a gene encoding a microsomal ω-3 fatty acid desaturase from *Arabidopsis thaliana*. Plant Physiol. 105: 767-768.

Prusky, D., Plumbley, R.A. and Kobiler, I. 1991. The relationship between antifungal

infection of unripe avocado fruits by *Colletotrichum* diene levels and fungal inhibition during quiescent *gloeosporioides*. Plant Pathol. 40:45-52.

Prusky, D. and Keen, N.T. 1995. Inducible preformed compounds and their involvement in the resistance of plants to pathogens. In R. Reuveni ed, Novel Approaches to Integrated Pest Management. Lewis Publishers, Boca Raton, pp. 139-152.

Sato, A., Becker, C.K., and Knauf, V. C. 1992. Nucleotide sequence of a complementary DNA clone encoding stearoyl-acyl carrier protein desaturase from *Simmondsia chinesis*. Plant Physiol. 99: 362-363.

Shanklin, J. and Somerville, C. 1992. Stearoyl-acyl-carrier desaturase from higher plants is structurally unrelated to the animal and fungal homolgs. Proc. Nat. Acad. USA 88:2410-2514.

Slocombe, S.P., Cummins, I., Jarvis, R.P., Murphy, D.J. 1992. Nucleotide sequence and temporal regulation of a seed-specific *Brassica napus* cDNA encoding a stearoyl-acyl carrier protein (ACP) desaturase. Plant. Mol. Biol. 20: 151-155.

Somerville, C. 1995. Direct tests of the role of membrane composition in low-temperature-induced photoinhibition and ch sensitivity plants and cyanobacreia. Proc. Natl. Acad. Sci USA 92: 6215- 6218.

Taylor, M.A., Smith, S.B., Davis, H.V. and Burch, L.R. 1992. The primary structure of a cDNA clone of stearoyl-acyl carrier protein desaturase gene from potato (*Solanum tuberosum* L.). Plant Physiol. 100: 533-534.

Thompson, G.A., Scherer, D.E., Foxall-Van Aken, S., Kenny, J.W., Young, H.L., Shintani, D.K., Kird, J.C. and Knauf, V.C. 1991. Primary structures of precursor and mature forms of stearoyl-acyl Carrier protein desaturase from safflower embryos and requirement of ferredoxin for enzyme activity. Proc. Natl. Acad. Sci. USA 88: 2578-2582.

Tzeng, D.D. and DeVay, J.E. 1993. Role of oxygen radicals in plant Disease development. Adv. Plant Pathol. 10:1-3.

Vick, B.A., Zimmermann, D.C. 1984. Biosynthesis of jasmonic acid by several plant species. Plant Physiol. 75: 458-46.

Wang, C., Chin, C.K. and Chen, A. 1999. Expression of the yeast Δ ⁹-desaturase gene in tomato enhances it resistance to powdery mildew. Physiol. Mol . Plant Pathol. 52: 371-383.

Yadav, N.S., Wierzbicki, A., Aegeter, M., Caster, C.S., P'erez-Grau, L., Kinney, A.J., Hitz, W.D., Booth, J.R., Schweiger, B., Ste, K.L., Allen, S.M., Blacwell, M.,

Reiter. R.S., Carlson, T. J., Russell, S.H., Feldmann, K.A., Piece, J. and Browse, J. 1993. Cloning of higher plant ω-3 fatty acid desaturase. Plant Physiol 103: 467-476.

Yakoby, N., Kobiler, I., Dinoor, A., Prusky, D. 2000. pH regulation of pectate lyase secretion modulates the attack of *Colletotrichum gloeosporioides* on avocado fruits. Appl. Environ. Microbiol. 66: 1026-1030.

Yong Moon, B., Higashi, S.I., Gombos, Z. and Murata, N. 1995. Unsaturation of the membrane lipids of chloroplasts stabilizes the photosynthetic machinery against low temperature photoinhibition in transgenic tobacco plants. Proc. Natl. Acad. USA 92: 6219-6223.

Legends to figures

Figure 1. Comparison of the deduced amino acid sequence of the avocado homologe (PAD1) encoding Δ^9 stearoyl-ACP desaturase and members of the Δ^9 -desaturase family. The entire coding sequence of PAD1 was compared with the *Ricinus communis* (castor), *Cucumis sativus* (cucumber), *Hlianthus annuus* and *brassica napus*. Identical or conserved residues are boxed.

Figure 2. Incorporation of C^{14} -linoleate into the antifungal diene in the presence of ethylene treated mesocarp fruit tissue.

Figure 3. Expression of PAD1 encoding Δ^9 stearoyl-ACP desaturase in response to exposure to CO₂, ethylene, wounding and inoculation with *C. gloeosporioides*. RNA was extracted (20 mg) from the mesocarp of fruits, electrophoresed and probed with *XhoI-EcoRI* fragment of PAD1, the avocado Δ^9 stearoyl-ACP desaturase. (EtBr-Ethidum bromide)

Figure 4. Expression of PAD1, Δ^9 stearoyl-ACP desaturase in response to light and low temperature. (A), Freshly harvested fruits were exposed for 6 h to dark and light

conditions at 4 0 C and 22 0 C. After the exposure period RNA was extracted (20 mg) from the mesocarp of fruits, electrophoresed and probed with *XhoI-EcoRI* fragment of PAD1, the avocado Δ^{9} stearoyl-ACP desaturase. (B) Quantification of hybridizing RNA is shown as percentage of maximum measured photostimulated luminescence (PSL). Correction for loading variation was done as described in Material and methods.

Figure 5. Expression of PAD1, Δ^9 stearoyl-ACP desaturase in fruits

pre-adapted to darkness, in response to light and low temperature (A), Freshly harvested fruits were adapted to darkness for 18 h at either 4 0 C or 22 0 C and then transferred to light for 6 h. After the exposure, total RNA was extracted from the mesocarp and subjected to Northern blot analysis (as described in Figure 2).

Figure 6. Linoleic acid composition and antifungal diene levels in mesocarp of fruit exposed for 6 h to dark at 4 0 C and 22 0 C.

Figure 7. (A), Decay development in fruits pre-exposed for 24 h to light at 22 0 C (\Diamond), Light at 4 0 C (\blacktriangle), Dark at 22 0 C (\blacksquare) or Dark at 4 0 C (x) and then artificially inoculated with *C. gloeosporioides*. (B), Decay development in naturally infected fruit pre-exposed to the same treatments as in A.

Table 1. Effect of exposure to ethylene, CO₂, wounding and inoculation on diene level

Diene level (mg/g.F.W.)

Pericarp	Mesocarp
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Control	646+35	1200+67
Ethylene*	1030+54 160	0+30
CO_2	1310+165	1834+145
Wounding	1150+30	1750+50
Inoculation	960+57	1647+65

^{*}The diene level as determined at 3 h after exposure ethylene, otherwise after 48 h.

3. PAPERS IN PREPARATION

Isolation of a cDNA Clone Encoding an Avocado ($Persea\ Americana$) Δ^{12} -Fatty Acid Desaturase (accession number: AY057406)

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The primary products of fatty acid synthesis are palmitoyl-acyl-carrier (16:0-ACP) and stearoyl-ACP (18:0-ACP). The soluble plastid enzyme, stearoyl-ACP desaturase (EC1.14.99.6), introduces the first double bond into stearoyl-ACP, between carbons 9 and 10, to produce oleoyl-ACP (18:1 Δ^9 -ACP). Additional double bonds are introduced into oleic acid or palmitic acid by Δ^{12} -desaturases after incorporation of the fatty acids into lipids (Shanklin and Somerville, 1991; Thompson et al., 1991). Oleate (18:1) produced by stearoyl-ACP Δ^9 -desaturase must be sequentially desaturated by Δ^{12} and Δ^{15} -desaturase to form the major fatty acid of photosynthetic tissues, α -linolenate (18:3) (Harwood, 1997). Microsomal Δ^{12} -desaturases transform oleic acid into linoleic acid preferably from exogenous oleoyl-CoA rather than elaidoyl-CoA (Lomascolo, et al., 1996).

It was hypothesised that Δ^{12} -fatty acid desaturases (EC1.14.99.-) may catalize the desaturation of the antifungal diene (Prusky and Keen, 1993; 1995; Prusky et al., 1991). Diene is enhanced by ethylene (Leikin-Frenkel and Prusky, 1998), CO₂, wounding or inoculation of avocado fruits with *C. Gloeosporioides* (Ardi et al., 1998).

Table 1. Characterization of Δ^{12} -desaturase cDNA from avocado (*Persea americana*)

Organism:

Avocado (Persea americana cv Fuerte).

Gene Product:

 Δ^{12} -desaturase (EC1.14.99.-).

Function:

Introduces the second double bond into oleic acid (18:1 $^{\Delta 9, \text{trans}}$) between carbon 12 and 13 to produce linoleic acid (18:2 $^{\Delta 9, 12}$).

Techniques:

Five different avocado genomic DNA fragments corresponding by sequence to Δ^{12} -desaturase were cloned by PCR with two degenerate primers, C A Y G A R T G Y G G N C A Y C A Y (upstream) and G C Y T T N K T N G C Y T C C A T (downstream) (synthesized by Life Technologies Ltd. UK), designed according to two boxes of conserved amino acid sequences, H E C G H H (upstream) and M E A t K A (downstream), of Δ^{12} -desaturases in higher plants. These fragments did not contain intron and were 692-bp long. One of the five DNA fragments served as probe to screen an avocado fruit cDNA library (constructed by L. Madi). An 867-bp cDNA clone with stop-codon and poly-A was obtained from the library. 5'-RACE PCR permitted cloning of the missed 5'-fragment, which contained 282-bp of cDNA with start-codon and 5-UTR sequences. Assembly of these two cDNAs generated a cDNA sequence containing the entire putative ORF of avocado Δ^{12} -desaturase with 3-UTR, poly-A and 36-bp 5-UTR.

Characteristics of cDNA:

The cDNA was 1250 bps long and encoded 382 amino in an open reading frame. Comparison of deduced amino acid sequence with *Arabidopsis thaliana* Δ^{12} -desaturase showed 83.8 % identity.

LITERATURE CITED

Ardi R, Kobiler I, Jacoby B, Keen NT Prusky D (1998) Involvement of epicatechin biosynthesis in the activation of the mechanism of resistance of avocado fruits to *Colletotrichum gloeosporioides*. Physiol Mol Plant Pathol **53:** 269-28

Harwood, JL (1997) Plant lipid metabolism. In: Plant Biochemistry, ed. P. M. Dey and J. B. Harborne. pp. 237-271. London: Academic Press

Leikin-Frenkel A, Prusky D (1998) Ethylene enhances the antifungal lipids contents

idioblasts from avocado mesocarp. Phytochemistry 49: 2291-2298

Lomascolo A, Dubreucq E, Galzy P (1996) Study of the Δ^{12} -desaturase system of *Lipomyces starkeyi*. Lipids **31:** 253-259

Prusky D, Keen NT (1993) Involvement of preformed antifungal compounds in the resistance of subtropical fruits to fungal decay. Plant Disease. **77:** 115-119

Prusky D, Keen NT (1995) Inducible preformed compounds and their involvement in the resistance of plants to pathogens. In: R. Reuveni ed, Novel Approaches to Integrated Pest Management. Lewis Publishers, Boca Raton, pp 139-152

Prusky D, Plumbley RA, Kobiler I (1991) The relationship between untifungal diene levels and fungal inhibition during quiescent infection of unripe avocado fruits by Colletotrichum gloesporioides. Plant Pathol **40:** 45-52

Shanklin J, Somerville C (1991) Stearoyl-acyl-carrier desaturase from higher plants is structurally unrelated to the animal and fungal homolgs. Proc Nat Acad Sci USA **88:** 2410-2514

Thompson GA, Scherer DE, Foxall-Van Aken S, Kenny JW, Young HL, Shintani DK, Kird1 JC, Knauf VC (1991) Primary structures of precursor and mature forms of stearoyl-acyl carrier protein desaturase from safflower embryos and requirement of ferredoxin for enzyme activity. Proc Natl Acad Sci USA 88: 2578-2582

Isolation of a cDNA Clone Encoding An Serine/threonine Kinase (accession number: AY057407) from Avocado Fruit (Persea americana)

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Enzymes of the eukaryotic protein kinase superfamily catalyze the reversible transfer of the γ -phosphate from ATP to amino acid side chains of proteins. In plants, protein phosphorylation has been implicated in responses to many signals, including light, pathogen invasion, hormones, temperature stress, and nutrient deprivation (Stone and Walker, 1995). Studies have implicated protein phosphorylation in the expression of disease resistance and self-incompatibility (Dietrich et al., 1990; Wang et al., 1996).

Serine/threonine kinases (EC 2.7.1.-) have been implicated as key elements in the signaling processes in plants (Sheen, 1996). They are found in various subcellular locations, which suggest that this family of kinases may be involved in multiple signaling pathways and the network of protein serine/threonine kinases in plant cells has been referred as a "central processor unit" (Hardie, 1999).

A fragment corresponding to serine/threonine kinase was cloned from an avocado substract cDNA library that was constructed under conditions of antifungal diene induction (Prusky and Keen, 1993; 1995; Prusky et al., 1991). The preliminary results indicted that this serine/threonine kinase might be induced under conditions (ethylene treatment) that induce antifungal diene synthesis.

Table 1. Characterization of serine/threonine kinase cDNA from avocado (*Persea americana*)

Organism:

Avocado (Persea americana cv Fuerte).

Gene Product:

Serine/threonine kinase (EC 2.7.1.-).

Function:

Serine/threonine kinase catalyze the reversible transfer of the γ -phosphate from ATP to serine/threonine amino acid side chains of proteins (Stone and Walker, 1995).

Techniques:

A 183-bp cDNA fragment corresponding to serine/threonine kinase was identified among cDNA clones from an avocado subtraction cDNA library that was constructed under conditions of antifungal diene induction. The preliminary results indicted that its mRNA transcription was induced under conditions that induce the antifungal diene synthesis. The DNA sequence information served to design primers for 3'-RACE PCR with 5'/3'-RACE PCR Kit following manufacturer's instruction, since the attempts to isolate the cDNA clone from avocado cDNA library failed. A total 337-bp sequence contained 165-bp DNA deducing the C-terminal protein of the putative protein, 3-UTR and poly-A was cloned by 3'-RACE PCR. An 896-bp fragment overlapping the 5-region of the 183-bp fragment was cloned from a mini genomic DNA library of avocado. No start codon and no any intron existed in this 896-bp fragment. A 5'-RACE PCR fragment of 206-bp long including 118-bp codoning region containing start codon that deducing N-terminal of the putative serine/threonine kinase protein, and 88-bp 5-UTR sequence was cloned. Assembly of these four fragments created a 1622-bp long of putative avocado serine/threonine kinase with 5-UTR, 3-UTR and poly-A tail.

Characteristics of cDNA:

The total size of the assembled serine/threonine kinase of avocado was 1622-bp long encoding a putative ORF of 453 amino acids and 88-bp of the 5-UTR and 172-bp of the 3-UTR including poly-A tail. The alignment of deduced 453 amino acid with serine/threonine kinase of *Arabidopsis thaliana* resulted 67.3% identity.

LITERATURE CITED

Dietrich A, Mayer J E and Hahlbrock K (1990) Fungal elicitor triggers rapid, transient, and specific protein phosphorylation in parsley cell suspension cultures. J. Biol. Chem. **265**: 6360-6368

Hardie DG (1999) Plant protein serine/threonine kinases: classification and functions. Annu. Rev. Plant Physiol. Plant Mol. Biol. **50:** 97-131

Prusky D, Keen NT (1993) Involvement of preformed antifungal compounds in the resistance of subtropical fruits to fungal decay. Plant Diease. **77:** 115-119

Prusky D, Keen NT (1995) Inducible preformed compounds and their involvement in the resistance of plants to pathogens. In: R. Reuveni ed, Novel Approaches to Integrated Pest Management. Lewis Publishers, Boca Raton, pp 139-152

Prusky D, Plumbley RA, Kobiler I (1991) The relationship between untifungal diene levels and fungal inhibition during quiescent infection of unripe avocado fruits by Colletotrichum gloesporioides. Plant Pathol **40:** 45-52

Sheen J (1996) Ca²⁺-dependent protein kinase and stress signal transduction in plants. Science **274**: 1900-1902

Stone JM, Walker JC (1995) Plant protein kinase families and signal transduction. Plant Physiology **108:** 451-457

Wang X, Zafian P, Choudhary M, and Lawton M (1996) The PR5K receptor protein kinase from *Arabidopsis thaliana* is structurally related to a family of plant defense proteins. Proc. Natl. Acad. Sci. USA **93:** 2598-2602.

Cloning and Expression of Fatty Acid Elongase from Avocado Related to Biosynthesis of Antifungal Diene

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Very-long-chain fatty acids (VLCFAs) have chain with 20 to 30 carbons, or more. The major site of VLCFA synthesis is the epidermal cells where they are utilized for the production of waxes embedded in the suberin and cutin and the epicuticular waxes which cover the aerial surfaces of the plant (Post-Beittenmiller, 1996). VLCFAs are synthesized by a microsomal fatty acid elongation (FAE) system by sequential additions of C2 moieties from malonyl coenzyme A (CoA) to pre-existing C18 fatty acids derived from the *de novo* fatty acid synthesis (FAS) pathway of the plastid (Fehling and Mukherjee, 1991; Fehling et al., 1992).

In *Arabidopsis*, mutations in the FAE1 gene result in highly reduced level of seed VLCFAs (James and Dooner 1990; Lemieux et al., 1990) and in a deficiency in acyl chain elongation activities from C₁₈ to C₂₂ and C₂₀ to C₂₂, suggesting that the product of the FAE1 gene is required for both elongation steps (Kunst et al., 1992). FAE1 is the rate-limiting enzyme for VLCFAs biosynthesis in *Arabidopsis* seed (Anthony and Ljerka, 1997).

It was hypothesized that fatty acid elongases may catalyze the carbon-chain elongation of preformed antifungal compound, 1-acetoxy-hydroxy-4-oxo-heneicosa-12, 15 diene in avocado fruits (*Persea americana*) (Prusky, 1996; Prusky and Keen, 1993; Prusky et al., 1982, 1983, 1985, 1991, 1994). Diene is enhanced by ethylene (Leikin-Frenkel and Prusky, 1998), CO₂, wounding or inoculation of avocado fruits with *C. gloeosporioides* (Ardi et al., 1998).

Cloning of Fatty Acid Elongase Genomic Fragments

Two avocado genomic DNA fragments of 226-bp long were cloned using the degenerative primers, CAYTTYTGYATHCAYGC (upstream) and SWRTTRCAYTTRAANCC (downstream). The primers were designed based in two conserved amino acids sequences HFCIHA (upstream) and GFKCNS (down stream) of KCS1 synthase from *Arabidopsis* (KCS1, AF053345), FAE1 KCS from *Arabidopsis* (FAE1, U29142), FAE KCS from *Brassica napus* (*B. napus*, U50771) and FAE KCS from jojoba (jojoba, U37088). These two genomic DNA sequences

code for 75 amino acids that differ only in 2 amino acids and no intron was found.

Based on the sequence information at their 5' end, we PCR cloned a 799-bp genomic DNA fragment from a mini genomic DNA library of avocado. This genomic sequence contained 441-bp exon at its 3' end and 358-bp intron at its 5' end.

Based on the same sequence information at of the 3' ends, we used 3'-RACE PCR and cloned a 333-bp fragment containing poly-A. It had 114-bp coding area and 219-bp 3'-UTR with poly-A.

Assembly of these three fragments created an 1141-bp sequence of the putative FAE of avocado. The amino acid alignment of the assembled sequence with FAE1 (U29142) of *Arabidopsis thaliana* shows high homology and 68.2% identity.

The expression of that gene during the elicitation of the antifungal diene is presently being studied.

Reference

Anthony AM and Ljerka K (1997) Very-long-chain fatty acid biosynthesis is controlled through the expression and specificity of the condensing enzyme. The Plant Journal, **12(1)**: 121-131

Ardi R, Kobiler I, Jacoby B, Keen NT, Prusky D (1998) Involvement of epicatechin biosynthesis in the activation of the mechanism of resistance of avocado fruits to *Colletotrichum gloeosporioides*. Physiol Mol Plant Pathol **53:**269-285

Fehling E, Lessire R, Cassagne C and Mukherjee KD (1992) Solubilization and partial purification of constituents of acyl-CoA elongase from *Lunaria annua*. Biochem. Biophys. Acta **1126**: 88-94

Fehling E and Mukherjee KD (1991) Acyl-CoA elongase from a higher plant (*Lunaria annua*): metabolic intermediates of very-long-chain acyl-CoA products and substrate specificity. Biochem. Biophys. Acta, **1082**: 239-246

James DW and Dooner HK (1990) Isolation of EMS-induced mutants in Arabidopsis altered in seed fatty acid composition. Theor. Appl. Genet. **80:** 241-245

Kunst L, Taylor DC and Underhill EW (1992) Fatty acid elongation in developing seeds of *Arabidopsis thaliana*. Plant Physiol. Biochem. **30:** 425-434

Leikin-Frenkel A and Prusky D (1998) Ethylene enhances the antifungal lipid content in idioblasts from avocado mesocarp. Phytochemistry **49:**2291-2298

Lemieux B, Miquel M, Somerville C and Browse J (1990) Mutants of *Arabidopsis* with alteration in seed lipid fatty acid composition. Theor. Appl. Genet. **80:** 234-240

Post-Beittenmiller D (1996) Biochemistry and molecular biology of wax production in plants. Annu. Rev. Plant physiol. Plant Mol. Biol. **47:** 405-430

Prusky D (1996) Pathogen quiescence in postharvest diease. Annu. Rev. Phytopathol. **34:**413-34

Prusky D and Keen NT (1993) Involvement of preformed antifungal compounds in the resistance of subtropical fruits to fungal decay. Plant Disease **77:**115-119

Prusky D, Keen NT and Eaks I (1983) Further evidence for the involvement of a preformed antifungal compound in the latency of *Colletottrichum gloeosporioides* on the unripe avocado fruits. Physiol. Plant Pathol. **22:**189-98

Prusky D, Keen NT, Sims JJ and Midland SL (1982) Possible involvement of an antifungal diene in the latency of *Colletottrichum gloeosporioides* on unripe avocado fruits. Phytopathology **72:**1578-1582

Prusky D, Kobiler I, Fishman Y, Sims JJ, Midland SL, Keen NT (1991) Identification of an antifungal compound in unripe avocado fruits and its possible involvement in the quiescent infection of *Colletotrichem gloeosporioides*. J. Phytopathol. **132**: 319-327

Prusky D, Kobiler I, Jacoby B, Sims JJ and Midland SL (1985) Effect of inhibitors of lipoxygenase activity and its possible relation to latency of *Colletottrichum gloeosporioides* on avocado fruits. Physiol. Mol. Pl. Path. **27:**269-279

Prusky D, Treeman S, Rodriguez RJ and Keen NT (1994) A nonpathogenic mutant strain of *Colletotrichum magna* induces resistance to *C. gloeosporioides* in avocado fruits. Molecular Plant-Microbe Interactions. **7:**326-333

Publication Summary (numbers)

	Joint	US	Israeli	Total
	IS/US	Authors	Authors	
	authorship	only	only	
Refereed (published, in press, accepted)	1		1	2
Submitted, in review, in preparation			3	3
Invited review papers				
Book chapters			4	4
Books				
Master theses				
Ph.D. theses			1	1
Abstracts			1	1

Not refereed (proceedings, reports, etc.)						
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Short Visits & Meetings	1	1	3	5		
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